Hydrogen-Bond Mediated Catalysis: The Aminolysis of 6-Chloropyrimidine as Catalyzed by Derivatives of Uracil

Kathryn N. Rankin, James W. Gauld, and Russell J. Boyd*

Contribution from the Department of Chemistry, Dalhousie University, Halifax, Nova Scotia, Canada B3H 4J3

Received October 31, 2000. Revised Manuscript Received January 4, 2001

Abstract: The aminolysis of 6-chloropyrimidine and 2-amino-6-chloropyrimidine has been examined by using density functional theory. Relative to the aminolysis of 6-chloropyrimidine, the addition of an electron-donating NH₂ group to C_2 increases the barrier to aminolysis, indicating that the third hydrogen bond does not play a catalytic role but introduces additional rigidity into the system. However, the computations suggest that there is an interesting correlation between the barrier to aminolysis and the proton affinity of the species that interacts with the incoming NH₃. To extend the range of proton affinities, the aminolysis of 6-chloropyrimidine was examined by using fluoro, imine, and thioketo derivatives of the uracil-derived bases. The proton affinity of the moiety that hydrogen bonds with NH₃ is decreased by fluoro substitution, and thus the aminolysis barriers are increased. Similarly, imine substitution enhances the PA of the moiety, which is reflected in a decrease in the aminolysis barriers. The same correlation exists for the thicketo-derived bases, whose PAs are intermediate between the fluoro and imine derivatives. Thus, the aminolysis of 6-chloropyrimidine and 2-amino-6-chloropyrimidine demonstrates the importance of a well-chosen proton acceptor and the catalytic possibilities associated with the formation of multiple hydrogen bonds.

Introduction

Hydrogen bonds are an essential feature of the structure and function of biological molecules. Although an individual hydrogen bond is relatively weak compared to a typical covalent bond, the cooperative nature of multiple hydrogen bonds confers stability to a complex,^{1,2} an important factor in the self-assembly of molecules.^{1,3,4} Due to the specificity of the donor—acceptor units and the inherent weakness of the individual bonds within a multiply hydrogen-bonded complex, molecules capable of forming hydrogen bonds have been employed as potential catalysts in reactions of organic and biological importance.^{3,5–8} Recently, nucleotide bases such as uracil have been utilized^{9,10} as catalytic agents due to the large variety of hydrogen bond functional groups associated with these molecules.¹¹

Tominaga et al.⁹ accelerated the aminolysis of 2-amino-6chloropurine by the addition of derivatives of uracil, which on

- * To whom correspondence should be addressed. E-mail: boyd@is.dal.ca. (1) Kolotuchin, S. V.; Zimmerman, S. C. J. Am. Chem. Soc. 1998, 120, 9092.
- (2) Beijer, F. H.; Sijbesma, R. P.; Vekemans, J. A. J. M.; Meijer, E. W.; Kooijman, H.; Spek, A. L. J. Org. Chem. **1996**, 61, 6371.
- (3) Kang, J.; Hilmersson, G.; Santamaria, J.; Rebek, J., Jr. J. Am. Chem. Soc. 1998, 120, 3650.
- (4) Sijbesma, R. P.; Beijer, F. H.; Brunsveld, L.; Folmer, B. J. B.; Hirschberg, J. H. K. K.; Lange, R. F. M.; Lowe, J. K. L.; Meijer, E. W. *Science* **1997**, 278, 1601.
- (5) Jubian, V.; Veronese, A.; Dixon, R. P.; Hamilton, A. D. Angew. Chem., Int. Ed. Engl. 1995, 34, 1237.
- (6) Jubian, V.; Dixon, R. P.; Hamilton, A. D. J. Am. Chem. Soc. 1992, 114, 1120.
 - (7) Kang, J.; Rebek, J., Jr. Nature 1997, 385, 50.
 - (8) Wang, B.; Sutherland, I. O. Chem. Commun. 1997, 1495.
- (9) Tominaga, M.; Konishi, K.; Aida, T. J. Am. Chem. Soc. 1999, 121, 7704.
- (10) Jorgensen, W. L.; Pranata, J. J. Am. Chem. Soc. 1990, 112, 2008.
 (11) Murray, T. J.; Zimmerman, S. C. J. Am. Chem. Soc. 1992, 114, 4010.

the basis of ¹H NMR evidence were assumed to form multiple hydrogen-bonding interactions. The latter were presumed to assist the formation of a reactive intermediate and to stabilize the transition state, giving rise to a catalytic enhancement in the observed rate of aminolysis. Subsequently, the present authors proposed a rationalization for the role of multiple hydrogen-bonding interactions on the basis of density functional theory calculations¹² carried out on a model reaction of the aminolysis of 6-chloropyrimidine. The uncatalyzed aminolysis was found to proceed with a sizable barrier, but through the addition of OCH₂, which forms a hydrogen bond to the incoming NH₃, the barrier to aminolysis was reduced. A further reduction in the barrier to aminolysis was obtained by enlarging the base to OHC-NH₂, which forms hydrogen bonds to both the incoming NH₃ and the N adjacent to the carbon at which substitution occurs.

Density functional theory is employed herein to investigate the role of the third hydrogen bond present in the aminolysis of 2-amino-6-chloropyrimidine (Scheme 1a). In addition, recent interest in the proton affinity of the proton donor^{13–15} involved in hydrogen bonding has prompted an investigation into the role of the hydrogen bond acceptor and the correlation between the proton affinity of the group that interacts with the incoming NH₃ moiety and the barrier to aminolysis. To examine this correlation, fluoro, imine, and thioketo derivatives of the uracilderived bases (OCH₂, OHC–NH₂, and OHC–NH–CHO) were utilized in the aminolysis of 6-chloropyrimidine (Scheme 1b).

⁽¹²⁾ Rankin, K. N.; Gauld, J. W.; Boyd, R. J. J. Am. Chem. Soc. 2000, 122, 5384.

⁽¹³⁾ Smith, D. M.; Golding, B. T.; Radom, L. J. Am. Chem. Soc. 1999, 121, 1383.

⁽¹⁴⁾ Smith, D. M.; Golding, B. T.; Radom, L. J. Am. Chem. Soc. 1999, 121, 5700.

⁽¹⁵⁾ Smith, D. M.; Golding, B. T.; Radom, L. J. Am. Chem. Soc. 1999, 121, 9388.





Computational Methods

Density functional theory calculations were carried out with use of the Gaussian 98^{16} suite of programs. Becke's three-parameter exchange functional (B3),^{17,18} as implemented¹⁹ in the Gaussian suite of programs, was used in conjunction with the correlation functional of Lee, Yang, and Parr (LYP).²⁰ Geometry optimizations were performed at the B3-LYP/6-31G(d,p) level. Harmonic vibrational frequencies and zero-point vibrational energy (ZPVE) corrections were calculated at the same level of theory. Relative energies were calculated at the B3-LYP/6-311+G-(2df,p) level by using the B3-LYP/6-31G(d,p) geometries and corrected with the appropriate ZPVE, i.e., B3-LYP/6-311+G(2df,p)//B3-LYP/6-31G(d,p) + ZPVE. The proton affinities for the uracil-derived bases were calculated at the aforementioned level of theory. All relative energies are in kJ mol⁻¹ and bond lengths in angstroms (Å). The optimized structures and total energies of all species are summarized in Tables S1 and S2, respectively, of the Supporting Information.

Results and Discussion

Previous calculations, utilizing OCH₂ and OHC-NH₂ to mimic the hydrogen-bonding functional groups in uracil alluded to the importance of hydrogen bonding to the incoming NH₃ moiety. While the isolated aminolysis was found to proceed with a sizable barrier of 138.1 kJ mol⁻¹, the barrier to aminolysis was reduced to 112.2 kJ mol⁻¹ by the addition of OCH₂ that formed an O····HNH₂ bond of 1.876 Å to NH₃. A further reduction in the aminolysis barrier to 95.3 kJ mol⁻¹ was attained by enlarging the base to OHC-NH₂ which formed two hydrogen bonds; a shorter O···HNH₂ bond of 1.766 Å to NH₃ and a longer NH ... N bond of 1.906 Å to the N adjacent to the carbon undergoing substitution. However, in the original study performed by Tominaga et al.,9 it is possible that the third hydrogen bond was involved in the aminolysis reaction. To assess the importance and function of the third hydrogen bond, an $-NH_2$ group was attached to C_2 of $Cl-C_4N_2H_3$ and the aminolysis of 2-amino-6-chloropyrimidine (Scheme 1a) was examined by using OCH₂, OHC–NH₂, OHC–NH–CHO, and 1-methyluracil as bases.

In the isolated aminolysis of 2-amino-6-chloropyrimdine (NH₃ + Cl-C₄N₃H₄), the reactants generate the initial complex **1a** (Figure 1a) lying 21.4 kJ mol⁻¹ lower in energy. As NH₃ remains hydrogen bonded to Cl-C₄N₃H₄, aminolysis proceeds via transition structure (TS) 1b with a sizable barrier of 158.8 kJ mol⁻¹. The addition of OCH₂ to NH₃ + Cl-C₄N₃H₄ (Figure 1b) forms complex 2a lying 5.1 kJ mol⁻¹ lower in energy. As OCH₂ remains bound to the incoming NH₃ moiety by a short O····HNH₂ bond of 1.888 Å, aminolysis proceeds via transition structure (TS) **2b** with a barrier of 117.8 kJ mol⁻¹. Enlarging the base to OHC-NH₂ generates the initial complex 3a lying 15.3 kJ mol⁻¹ lower in energy upon addition to $NH_3 + Cl$ -C₄N₃H₄ (Figure 1c). Aminolysis of Cl-C₄N₃H₄ proceeds via TS **3b**, a barrier of 99.9 kJ mol⁻¹, in which OHC-NH₂ generates a noticeably shorter O····HNH₂ bond of 1.797 Å to the incoming NH₃ and forms an N····HN bond (1.930 Å) to N_1 of Cl-C₄N₃H₄. With OHC-NH-CHO as the base (Figure 1d), complex 4a is generated lying 34.4 kJ mol⁻¹ lower in energy than NH₃ + Cl-C₄N₃H₄. As OHC-NH-CHO remains bound to NH₃ via an elongated O····HNH₂ bond (1.869 Å) and bound to $Cl-C_4N_3H_4$ by a shorter N···HN bond (1.725 Å) to N_1 , aminolysis proceeds via TS 4b with a noticeable increase in the barrier to 110.3 kJ mol⁻¹. Finally, the addition of 1-methyluracil to 2-amino-6-chloropyrimidine (Figure 1e) generates the initial complex **5a** lying 24.7 kJ mol^{-1} lower in energy. Aminolysis proceeds via TS 5b with a barrier of 103.6 kJ mol⁻¹ in which 1-methyluracil forms a short O····HNH₂ bond (1.809 Å) to NH₃, an elongated N····HN bond (1.792 Å) to N₁ of Cl- $C_4N_3H_4$, and an O···HNH bond (2.025 Å) with the amino group at C₂ of Cl-C₄N₃H₄. For the purpose of comparison, the addition of OCH-NH-HCO (Figure S2a) and 1-methyluracil (Figure S2b) to 6-chloropyrimidine $(NH_3 + Cl - C_4N_2H_3)$ yields barriers to aminolysis of 100.8 and 95.2 kJ mol⁻¹, respectively.

The barriers to aminolysis for 2-amino-6-chloropyrimidine (Scheme 1a) and 6-chloropyrimidine (Scheme 1b) are summarized in Table 1. Relative to the aminolysis of 6-chloropyrimidine, the presence of the amino group in 2-amino-6chloropyrimidine increases the barriers by 5.6 and 4.6 kJ mol⁻¹, for OCH₂ and OHC-NH₂, respectively. The presence of the electron-donating NH₂ group results in C₆, the carbon undergoing substitution, being less susceptible to nucleophilic attack by the incoming NH₃, and as a consequence, the C···Cl distance in the transition structures is elongated by ~ 0.02 Å. Enlarging the model base to OHC-NH-CHO, which forms a third hydrogen bond in the aminolysis of 2-amino-6-chloropyrimidine, leads to a barrier 9.5 kJ mol⁻¹ larger than that observed in the aminolysis of 6-chloropyrimidine. Due to the formation of the third hydrogen bond, the electron donating ability of $-NH_2$ is further enhanced, resulting in an additional decrease in the electrophilicity of the C undergoing substitution. The addition of 1-methyluracil to 2-amino-6-chloropyrimidine exhibits a similar effect with a barrier to aminolysis that is 8.4 kJ mol⁻¹ larger than that observed for 6-chloropyrimidine. Thus, the absence of a decrease in the barrier to aminolysis for 2-amino-6-chloropyrimidine and the minor changes in the transition structure geometry indicate that the third hydrogen bond does not play a catalytic role in the aminolysis reaction. However, the partial double bond character of the C-NH₂ bond provides a more rigid framework upon which the aminolysis reactions may proceed.

Closer examination of the aminolysis of 6-chloropyrimidine reveals a correlation between the barriers to aminolysis and the

⁽¹⁶⁾ Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Zakrzewski, V. G.; Montgomery, J. A.; Stratmann, R. E.; Burant, J. C.; Dapprich, S.; Millam, J. M.; Daniels, A. D.; Kudin, K. N.; Strain, M. C.; Farkas, O.; Tomasi, J.; Barone, V.; Cossi, M.; Cammi, R.; Mennucci, B.; Pomelli, C.; Adamo, C.; Clifford, S.; Ochterski, J.; Petersson, G. A.; Ayala, P. Y.; Cui, Q.; Morokuma, K.; Malick, D. K.; Rabuck, A. D.; Raghavachari, K.; Foresman, J. B.; Cioslowski, J.; Ortiz, J. V.; Stefanov, B. B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Gomperts, R.; Martin, R. L.; Fox, D. J.; Keith, T.; Al-Laham, M. A.; Peng, C. Y.; Nanayakkara, A.; Gonzalez, C.; Challacombe, M.; Gill, P. M. W.; Johnson, B. G.; Chen, W.; Wong, M. W.; Andres, J. L.; Head-Gordon, M.; PA, 1998.

⁽¹⁷⁾ Becke, A. D. J. Chem. Phys. 1993, 98, 5648.

⁽¹⁸⁾ Becke, A. D. J. Chem. Phys. 1993, 98, 1372

⁽¹⁹⁾ Stephens, P. J.; Devlin, F. J.; Chabalowski, C. F.; Frisch, M. J. J. Phys. Chem. **1994**, 98, 11623.

⁽²⁰⁾ Lee, C.; Wang, W.; Parr, R. G. Phys. Rev. B 1987, 37, 785.



Figure 1. (a) Schematic energy profile for the aminolysis of 2-amino-6-chloropyrimidine with (b) OCH_2 hydrogen bonded to the incoming NH_3 moiety (see text). (c) $OHC-NH_2$ hydrogen bonded to both the incoming NH_3 moiety and the pyrimidine ring (see text). (d) OHC-NH-CHO hydrogen bonded to both the incoming NH_3 moiety and the pyrimidine ring (see text). (e) 1-Methyluracil hydrogen bonded to both the incoming NH_3 moiety and the pyrimidine ring (see text).

O···HNH₂ hydrogen bond distance, a consequence of the proton affinity (PA) of the carbonyl group of the base involved in hydrogen bonding to NH₃. In Table 2, the proton affinities of the uracil-derived bases are summarized. For the aminolysis of 6-chloropyrimidine, the barrier to aminolysis decreases from 112.2 kJ mol⁻¹ for OCH₂ to 100.8 and 95.3 kJ mol⁻¹ for the addition of OHC–NH–CHO and OHC–NH₂, respectively. For these three bases, the decrease in the barrier to aminolysis of 6-chloropyrimidine is correlated with an increase in the PA of the terminal carbonyl group interacting with NH₃, as is evident by the decrease in the O···HNH₂ distance from 1.876 Å¹² to

1.839 and 1.766 Å.¹² As expected, a further decrease in the barrier to aminolysis is attained by the use of 1-methyluracil, which has the largest PA of the bases examined. However, there is a slight increase in the O···HNH₂ distance (1.787 Å), a consequence of the electron-donating influence of the methyl group.

The same qualitative trends are evident for the aminolysis of 2-amino-6-chloropyrimidine. The barrier to aminolysis (and the O···HNH₂ distances) decreases from 117.8 kJ mol⁻¹ (1.888 Å) for OCH₂ to 110.3 kJ mol⁻¹ (1.869 Å) and 99.9 kJ mol⁻¹ (1.797 Å) for OHC–NH–CHO and OHC–NH₂, respectively. The

 Table 1.
 Summary of the Barriers to Aminolysis (kJ mol⁻¹)

 Involving the Uracil-Derived Bases

	$NH_3 + Cl - C_4N_3H_4$	$NH_3 + Cl - C_4N_2H_3$		
base	X = 0	X = O	X = S	X = NH
uncatalyzed	158.8	138.1		
XCH ₂	117.8	112.2	117.8	107.6
XHC-NH ₂	99.9	95.3	102.6	90.6
XHC-NH-CHHO	110.3	100.8	106.9	98.3
1-methyluracil	103.6	95.2		
XCHF		117.1		
$\mathbf{X}FC-NH_2$		100.9		
XFC-NH-CHO		105.3		

Table 2. Calculated^{*a*} Proton Affinities (kJ mol⁻¹) of the Carbonyl Oxygens in the Uracil-Derived Bases

	proton affinities		
base	X = O	X = S	X = NH
XCH ₂	700.9	763.6	861.9
XHC-NH-CHO	789.9	801.8	896.2
$XHC-NH_2$	808.5	850.6	945.8
1-methyluracil ^b	861.5		
1-methyluracil ^c	831.2		
XCHF	646.8		
XFC-NH-CHO	701.1		
$\mathbf{X}FC-NH_2$	743.7		

 a See theoretical methods. b Carbonyl oxygen that interacts with the incoming NH₃. c Carbonyl oxygen that interacts with $-NH_{2}$ on 2-amino-6-chloropyrimidine.

addition of 1-methyluracil to 2-amino-6-chloropyrimidine marginally increases the aminolysis barrier by 3.7 kJ mol⁻¹ (103.6 kJ mol⁻¹), whereas there is no effect for the corresponding aminolysis of 6-chloropyrimidine. In this instance, the PA of the carbonyl group of 1-methyluracil interacting with -NH₂ of 2-amino-6-chloropyrimidine is larger than that associated with the corresponding group in OHC-NH-CHO (Table 2). Thus, utilizing 1-methyluracil as the base in the aminolysis enhances the electron density in the pyrimidine ring to a greater degree than that observed when OHC-NH-CHO acts as the base. While the carbonyl group in 1-methyluracil (which interacts with NH₃) has the largest PA of the molecules examined, it is insufficient to compensate for the decreased electrophilicity of the C at which substitution occurs. Thus, while the general trend in barriers to aminolysis is consistent with the PA of the terminal group interacting with NH₃, the electronic effect associated with the formation of the third hydrogen bond in the aminolysis of 2-amino-6-chloropyrimidine must also be considered.

Thus, the barriers to aminolysis of 6-chloropyrimidine correlate with the PA of the carbonyl oxygen of the base that hydrogen bonds with the incoming NH₃ moiety. This correlation is also prevalent for the aminolysis of 2-amino-6-chloropyrimidine although geometrical factors must be considered. Since the formation of the third hydrogen bond does not catalyze the aminolysis reaction, the model reaction of 6-chloropyrimidine was employed to further examine the correlation between the PA of the portion of the base that hydrogen bonds to the incoming NH₃ and the calculated barrier to aminolysis. To provide a range of PAs, derivatives of the carbonyl bases were examined in which the PA of the carbonyl-derived group was modified by fluorine substitution, which decreases the PA of the carbonyl oxygen, and by imine substitution, which leads to a larger PA.

Fluorine Substitution. Replacement of the H adjacent to the carbonyl group of the three smallest bases involved in hydrogen bonding to the NH₃ moiety by an electron-withdrawing fluorine yields OCHF, OFC–NH₂, and OFC–NH–CHO. Addition of



Figure 2. Schematic energy profile for the aminolysis of 6-chloropyrimidine with (a) OCHF hydrogen bonded to the incoming NH_3 moiety (see text), (b) OFC $-NH_2$ hydrogen bonded to both the incoming NH_3 moiety and the pyrimidine ring (see text), and (c) OFC-NH-CHO hydrogen bonded to both the incoming NH_3 moiety and the pyrimidine ring (see text).

OCHF to NH₃ + Cl-C₄N₂H₃ (Figure 2a) generates complex **6a** lying 14.2 kJ mol⁻¹ lower in energy. Aminolysis proceeds via TS **6b** with a barrier of 117.1 kJ mol⁻¹ and a long O··· HNH₂ bond of 1.936 Å. With OFC-NH₂ as the base, complex **7a** is generated lying 27.4 kJ mol⁻¹ lower in energy upon addition to NH₃ + Cl-C₄N₂H₃ (Figure 2b). Aminolysis proceeds via TS **7b** with a reduced barrier of 100.9 kJ mol⁻¹ and a significantly shorter O···HNH₂ bond of 1.816 Å. Addition of OFC-NH-CHO to NH₃ + Cl-C₄N₂H₃ (Figure 2c) generates complex **8a** lying 29.1 kJ mol⁻¹ lower in energy. Ami-

nolysis proceeds via TS **8b** with a barrier of 105.3 kJ mol⁻¹ and an O····HNH₂ bond of 1.888 Å.

For the fluorine-derived bases, the barrier to aminolysis of 6-chloropyrimidine decreases in the sequence OCHF, OFC– NH–CHO, and OFC–NH₂. As summarized in Table 2, this decrease in barrier height is associated with the increase in the PA of the terminal oxygen that interacts with NH₃. As observed for the uracil-derived bases, increasing the PA of the terminal oxygen shortens the O···HNH₂ distance of the transition structures from 1.936 Å for OCHF to 1.888 Å for OFC–NH–CHO and to 1.816 Å for OFC–NH₂. As the PA of the fluoro-substituted uracil bases is lower than that of unsubstituted uracil-derived bases, it follows that the barrier to aminolysis is larger by ~5 kJ mol⁻¹ and the O···HNH₂ distances are longer by ~0.05 Å.

Imine Substitution. Due to the abundance of nitrogencontaining species in biological systems and the evidence that the nitrogen in the imine group is a better proton acceptor than the carbonyl oxygen,²¹ and hence has a larger PA, the aminolysis of Scheme 1b was reexamined with the carbonyl oxygen replaced by an imine group to produce the bases HNCH₂, HNCH-NH₂, and HNCH-NH-HCO. The addition of HNCH₂ to $NH_3 + Cl - C_4N_2H_3$ (Figure 3a) generates complex **9a** lying 10.7 kJ mol⁻¹ lower in energy. As HNCH₂ remains bound to NH₃ by a short N····HNH₂ bond of 1.878 Å, aminolysis proceeds via TS 9b with a barrier of 107.6 kJ mol⁻¹. Utilizing HNCH- NH_2 as the base in the aminolysis of $NH_3 + Cl - C_4N_2H_3$ (Figure 3b) produces complex **10a** lying 21.5 kJ mol⁻¹ lower in energy. Aminolysis proceeds via TS 10b with a barrier of 90.6 kJ mol⁻¹ and a shorter N····HNH₂ bond of 1.800 Å. Finally, the addition of HNCH-NH-HCO to $NH_3 + Cl-C_4N_2H_3$ (Figure 3c) produces complex **11a** lying 22.5 kJ mol⁻¹ lower in energy. Aminolysis of $NH_3 + Cl - C_4N_2H_3$ proceeds with a barrier of 98.3 kJ mol⁻¹ and an N····HNH₂ distance of 1.875 Å in TS 11b.

Thus, the barrier to aminolysis is reduced from $107.6 \text{ kJ mol}^{-1}$ when HNCH₂ is used as the base in the reaction of 6-chloropyrimidine to 98.3 and 90.6 kJ mol⁻¹ with NHCH–NH–HCO and HNCH–NH₂, respectively. As the barrier decreases, the N···HNH₂ distance shortens from 1.878 Å with HNCH₂ as the base to 1.875 and 1.800 Å with NHCH–NH–HCO and HNCH–NH₂, respectively. The shortening of the N···HNH₂ distances is consistent with a sequential increase in the PA associated with the imine that interacts with NH₃ (Table 2). This is the same trend as that observed for the uracil- and fluorine-derived bases examined in the aminolysis of 6-chloropyrimidine and provides a range of behavior between the small PAs associated with the fluorine-derived bases and the high PAs of the imine-derived bases.

Sulfur Substitution. Sulfur may act as a hydrogen bond acceptor and is known to replace oxygen in this function. However, due to its larger size, sulfur is expected to act as a weaker hydrogen bond acceptor than oxygen. To determine if the correlation between the PA of the base that interacts with NH₃ and the barrier to aminolysis is maintained as the carbonyloxygen of the base is replaced by sulfur, the aminolysis of 6-chloropyrimidine (Scheme 1b) was examined with SCH₂, SCH–NH₂, and SCH–NH–HCO as bases.

The aminolysis of 6-chloropyrimidine with SCH₂ as the base proceeds via TS **12b** (Figure S2a) with a barrier of 117.8 kJ mol⁻¹ and an S···HNH₂ bond of 2.442 Å. The PA of the thicketo group is increased upon enlarging the base to SCH–



Figure 3. Schematic energy profile for the aminolysis of 6-chloropyrimidine with (a) $HNCH_2$ hydrogen bonded to the incoming NH_3 moiety (see text), (b) $HNCH-NH_2$ hydrogen bonded to both the incoming NH_3 moiety and the pyrimidine ring (see text), and (c) HNCH-NH-CHO hydrogen bonded to both the incoming NH_3 moiety and the pyrimidine ring (see text).

NH₂ (Figure S2b) and the aminolysis proceeds via TS **13b** with a notably reduced barrier of 102.6 kJ mol⁻¹ and a shortened S···HNH₂ bond (2.335 Å). As the base is enlarged further to SCH-NH-HCO, which has a lower PA than that observed for SCH-NH₂, the aminolysis of 6-chloropyrimdine (Figure S2c) proceeds via TS **14b** with a slightly larger barrier of 106.9 kJ mol⁻¹ and an elongation of the S···HNH₂ bond by 0.053 Å (2.388 Å). While the PAs of the thioketo bases (Table 2) are intermediate between those of the carbonyl- and the imine-

⁽²¹⁾ Scheiner, S. Hydrogen Bonding: A Theoretical Perspective; Oxford University Press: New York, 1997.

derived bases, the barriers are slightly larger than those reported for the aminolysis of 6-chloropyrimidine employing OCH₂, OCH-NH₂, and OCH-NH-HCO as bases. The larger size of the sulfur atom results in an elongation of the S···HNH₂ bond to the incoming NH₃, and as such, hydrogen bonding does not enhance the electron-donating ability of NH₃ to the same degree as was observed for the uracil-derived bases. Nonetheless, the correlation between the PA of the terminal thioketo group that hydrogen bonds to NH₃ and the barrier heights is evident.

Conclusions

The aminolysis of 6-chloropyrimidine and 2-amino-6-chloropyrimdine has been investigated by density functional theory calculations. Comparison of the barriers for aminolysis of 6-chloropyrimidine to those calculated for the aminolysis of 2-amino-6-chloropyrimidine reveals that the presence of the $-NH_2$ group enhances the electron density in the pyrimidine ring which in turn diminishes the electrophilicity associated with the C at which substitution occurs. While the formation of the third hydrogen bond does not act as a catalyst in the reaction, it does provide a more rigid skeleton upon which the aminolysis reaction may proceed.

Closer examination of the aforementioned aminolysis reactions has revealed a correlation between the observed barrier and the proton affinity of the carbonyl group of the base that forms the O···HNH₂ hydrogen bond to the incoming NH₃ group. To further investigate the correlation between the PA of the base interacting with the NH₃ and the barrier to aminolysis, the chemical nature of the proton acceptor, i.e., the base, was altered. Replacement of the H adjacent to the carbonyl group by a fluorine atom decreases the calculated PA of the proton acceptor relative to that observed for the nonsubstituted bases and, hence, increases the barrier to aminolysis of 6-chloropyrimidine. Similarly, the imine derivatives of the carbonyl bases have larger PAs than the carbonyl-derived bases and therefore decrease the barrier to aminolysis. While the results for the fluorine- and imine-derived bases provide evidence for the important role of the proton acceptor, thioketo substitution was also examined to extend the scope of the study. Although the sulfur-derived bases possess a proton affinity intermediate between the carbonyl-and fluoro-derived bases, they generate elongated bonds to NH₃ which is reflected in the aminolysis barrier of 6-chloropyrimidine being slightly larger than that observed for the uracil-derived species.

Thus, the aminolysis of 6-chloropyrimidine and 2-amino-6chloropyrimidine illustrates the ability of the functional groups in uracil to catalyze the reaction by the formation of multiple hydrogen bonds, which stabilize the transition structures. Thus the aminolysis reaction provides a clear example of the catalytic possibilities associated with the formation of multiple hydrogen bonds and illustrates the importance and flexibility associated with a well-chosen hydrogen bond acceptor.

Acknowledgment. We gratefully acknowledge the Natural Sciences and Engineering Research Council of Canada (NSERC) and the Killam Trusts for financial support.

Supporting Information Available: Archive entries of the B3-LYP/6-31G(d,p) optimized structures (Table S1), total electronic energies of all species in the study (Table S2), schematic energy profiles for the aminolysis of 6-chloropyrimidine with OCH–NH–HCO and 1-methyluracil as bases (Figure S1, parts a and b, respectively), schematic energy profiles for the sulfur-derived bases (Figure S2a–c) and charges on the heavy atoms in the various transition structures from Mulliken population analyses (Figure S3) (PDF). This material is available free of charge via the Internet at http://pubs.acs.org.

JA0038373